



ELSEVIER

Journal of Hepatology 40 (2004) 341–352

Journal of  
Hepatology

[www.elsevier.com/locate/jhep](http://www.elsevier.com/locate/jhep)

Review

## Extrahepatic disease manifestations of HCV infection: some current issues

Vincent Agnello<sup>1,2,\*</sup>, Francesco G. De Rosa<sup>1,†</sup>

<sup>1</sup>Department of Laboratory Medicine, Lahey Clinic Medical Center, 41 Mall Road, Burlington, MA 01805, USA

<sup>2</sup>Edith Norse Rogers Memorial Veterans Affairs Hospital, Bedford, MA, USA

### 1. Introduction

Productive hepatitis C virus (HCV) infection appears to be primarily, and in most patients almost exclusively, confined to the liver. However, a wide variety of extrahepatic disease manifestations have been reported to be associated with the infection. The prevalence of extrahepatic disease manifestations is not known with certainty but these observations have suggested that HCV is involved in non-hepatic pathological processes.

There are two immunologic features of HCV infection that may predispose to extrahepatic disease manifestations. The first feature is the virus avoids immune elimination. This feature leads to chronic infection, accumulation of circulating immune complexes, and autoimmune phenomena associated with chronic viral infections. The second feature is the virus stimulates production of monoclonal rheumatoid factors (mRF). In the majority of patents mRF is highly restricted and has been dubbed 'WA' mRF. This feature causes the type II cryoglobulinemia associated with HCV infection (HCV-type II cryoglobulinemia) that is responsible for most of the symptomatic cryoglobulinemic vasculitis, or Melzer-Franklin syndrome. Although this manifestation occurs relatively infrequently, as do all the extrahepatic disease manifestations, it is the major extrahepatic disease manifestation of HCV infection due to the increased morbidity and mortality accompanying the disease.

A widely postulated but controversial hypothesis is that extrahepatic disease manifestations are caused by extrahepatic tropisms of HCV, particularly lymphotropism. The latter is thought to be involved in the production of autoantibodies and non-Hodgkin's B-cell lymphoma (B-NHL). In this review, the emphasis will be on some of

these topics as well as other current issues. Only a brief overview of all the extrahepatic disease manifestations associated with HCV infection will be given. A comprehensive review of the subject has been published [1].

### 2. Extrahepatic disease manifestations associated with HCV infection

At least 36 extrahepatic disease manifestations, mainly autoimmune disorders, have been reported to be associated with HCV infection (Table 1) [2–69]. There are conflicting reports for most of these associations and definitive validation is lacking for all except the association with mixed cryoglobulinemia. In most reports relatively small numbers of patients and an inadequate numbers of control subjects were studied. In many studies there was a problem with patient selection bias that can produce false associations especially in endemic regions where coincidental disease is more likely to be observed. The true prevalence of any of these manifestations is not known because studies have not been performed on large numbers of unselected HCV-infected patient populations. In all cases, with the exception of symptomatic mixed cryoglobulinemia, the critical determination of a specific role for HCV in the pathogenesis has not been established.

Despite the limitations of the studies noted above, observations on extrahepatic disease manifestations have been taken as evidence that HCV has tropism for cells other than hepatocytes, particularly lymphocytes. It has been postulated that HCV infection of lymphocytes is the cause of the increased autoimmune disease reported to be associated with HCV infection. However clinical observations dispute this hypothesis. The autoantibodies in patients with HCV infection appear to resemble the physiologic type accompanying many viral infections. The autoantibody titers are low, there is no female gender predominance, there is no association with HLA-DR genes, and there are usually no clinical autoimmune

\* Corresponding author. Tel.: +1-781-273-8887; fax: +1-781-744-5208.

E-mail address: [vincent.agnello@lahey.org](mailto:vincent.agnello@lahey.org) (V. Agnello).

† Present address: Department of Infectious Diseases, University of Turin, Turin, Italy.

**Table 1**  
**Reported extrahepatic disease associations with HCV infection [2–69]**

Antiphospholipid syndrome	MALToMa
Aplastic anemia	Membranoproliferative glomerulonephritis
Autoimmune hemolytic anemia	Membranous glomerulonephritis
Autoimmune thyroiditis	Mixed cryoglobulinemia
Chronic fatigue syndrome	Mooren corneal ulcers
Behcet's syndrome	Multiple myeloma
Carotid atherosclerosis	Non-Hodgkin's lymphoma
CRST syndrome	Neurocognitive Impairment
Dermatomyositis	Pancreatitis
Diabetes mellitus	Polyarteritis nodosa
Fibromyalgia	Polymyositis
Guillain-Barré syndrome	Porphyria cutanea tarda
Hypertrophic cardiomyopathy	Rheumatoid arthritis
Hypocholesterolemia	Sialadenitis
Idiopathic pulmonary fibrosis	Sjögren's syndrome
Idiopathic thrombocytopenia purpura	Systemic lupus erythematosus
IgA deficiency	Uveitis
Lichen planus	Waldenstrom's macroglobulinemia

manifestations. For example, the anticardiolipin antibodies in HCV infection occur in low titers, are not associated with thrombocytopenia or thrombotic events, and the anti  $\beta_2$ -glycoprotein antibodies usually associated with clinical manifestations are not present [2,3]. This difference is also illustrated by the fact that autoantibodies associated with HCV infection are distinct from those found in autoimmune hepatitis, an established clinical autoimmune disease [70].

For most autoantibodies that occur in HCV-infected patients, the prevalence and significance are in question. In contrast, it is has been widely documented that there is an increased prevalence of polyclonal rheumatoid factors (RF). Production of RF occurs in response to circulating complexes of IgG and occurs in high prevalence predominantly in patients with prolonged chronic infection. Hence, the mechanism is distinct from that involved in the production of other autoantibodies. Production of mRF also appears to be distinct from both the production of RF and other autoantibodies. On B cell clonal analysis mRF sequences are detectable in HCV-infected patients without cryoglobulinemia and do not correlate with the presence of RF [71]. Also consistent with a distinct mechanism for the production of mRF is that the prevalence of other autoantibodies in type II cryoglobulinemia is low [72].

Several associations of extrahepatic disease manifestations with HCV infection other than symptomatic type II cryoglobulinemia are notable. There appears to be a strong association of salivary gland lesions, lymphocytic capillaritis, and lymphocytic adenitis with chronic hepatitis C [73]. The clinical characteristics of patients with these lesions, however, differed from those of patients with Sjögren's syndrome by a predominantly male rather than female sex prevalence, absence of serum antinuclear antibodies, low rather than high association with HLA DR 3, milder histopathology (predominantly grades I–II rather than grades II–IV), and clinical symptoms (only 30% had

xerostomia and none had xerophthalmia). Despite several reports of increased prevalence of HCV in patients with Sjögren's syndrome it is clear that the virus is neither necessary nor sufficient to produce this disease [74].

A study of 48 patients who had Sjögren's syndrome without mixed cryoglobulinemia were all positive for SSA/Ro but none had HCV infection [75]. HCV infection is common only in patients with Sjögren's syndrome associated with symptomatic type II cryoglobulinemia. However, the prevalence of Sjögren's syndrome in type II cryoglobulinemia associated with HCV infection is only 6% [1]. There are conflicting results from studies of salivary tissue from patients with HCV infection [76,77]. In HCV infected patients with sialadenitis or Sjögren's syndrome, HCV was detected in epithelial cells of salivary gland but there were no differences in the lesions in HCV infected and non-infected patients [76]. Studies on salivary glands from HCV infection without salivary gland abnormalities were negative [77]. Similarly, in palpable purpura lesions in patients with HCV-type II cryoglobulinemia, HCV was detected in keratinocytes and glandular epithelial cells in the skin in the inflammatory lesions but not in normal skin. Up-regulation of LDL receptors was demonstrated on keratinocytes in the lesional but not the normal skin [78]. Hence, up-regulation of LDL receptors associated with inflammation may result in uptake of HCV since the LDL receptor has been shown to mediate endocytosis of the virus [79].

Of interest in regard to potential mechanisms involved in sialadenitis in HCV infection is that studies in transgenic mice expressing HCV envelope genes found sialadenitis that had features similar to the lesions in humans, i.e. no gender prevalence and negative antinuclear autoantibodies [80]. Despite the non-physiologic over-expression of envelope proteins in this animal model [81], the results suggest that the HCV virus may be directly involved in sialadenitis rather than triggering an autoimmune process. The finding in this mouse model may also relate to the occurrence of lichen planus in HCV infection. As with Sjögren's syndrome it is clear that the HCV is neither sufficient nor necessary for production of lichen planus [74] and a recent study has questioned the increased prevalence of lichen planus among HCV-infected patients that has been reported in endemic areas [82]. However it has been demonstrated that in patients with lichen planus, HCV specific CD4 + and CD8 + lymphocytes are detectable in the lichen planus lesions but not in the blood [83]. Hence, in both Sjögren's syndrome and lichen planus HCV may be a contributing but not a primary pathologic factor.

Very high prevalence of HCV infection has been reported among patients with porphyria cutanea tarda in Southern Europe and Japan but not in Northern Europe and Australia suggesting again a possible patient selection bias in endemic regions [84]. Yet a high prevalence was also found in the United States and it has been suggested that HCV is a precipitating factor for porphyria cutanea tarda [85]. The mechanism whereby HCV would precipitate

the disease is unclear since the infection does not decrease activity of hepatic uroporphyrinogen decarboxylase, which is deficient in active porphyria cutanea tarda [86]. Confounding these studies is the high prevalence of alcoholism among patients with porphyria cutanea tarda since alcohol is a known risk factor for precipitating the disease. Therefore, it remains questionable whether HCV alone can trigger porphyria cutanea tarda.

### 3. Current issues

#### 3.1. Prevalence and clinical significance of mixed cryoglobulinemia

Meltzer et al. described clinical manifestations associated with ‘mixed’ cryoglobulinemia in 1966. They described a ‘syndrome’ that consisted of the triad of palpable purpura, arthralgias and weakness plus other clinical features, such as glomerulonephritis, lymphadenopathy, splenomegaly, and hepatomegaly in some of the patients [87]. Cryoglobulinemia in these patients had a ‘mixed’ composition of IgG and IgM RF. Studies performed by the Franklin group, as well as others, implicated the cryoglobulin components in the immune complex mediated vascular and glomerular lesions that occurred in many of these patients. It is now apparent that the triad of palpable purpura, arthralgias and weakness occurs in a minority of patients and that the main characteristic of the disease is the markedly heterogeneous manifestations of a systemic vasculitis with palpable purpura being the most prevalent.

The later classification of cryoglobulins based on immunoglobulin (Ig) composition by Brouet and colleagues [88], which defined three types of cryoglobulins, was also based on the characterization of the cryoglobulins from patients with disease manifestations. Type I consisted of a single monoclonal Ig; types II and III, or mixed cryoglobulins, consisted of polyclonal IgG and monoclonal (type II) or polyclonal (type III) IgM with RF activity. Because RF was a component of most of the types II and III cryoglobulins, the original Brouet study was essentially a correlation of IgG-IgM RF complexes with manifestations of immune complex diseases, as were the studies by the Franklin group. In recent years there has been widespread study of HCV infected patients without extrahepatic clinical manifestations.

The prevalence of mixed cryoglobulinemia increases with the duration of the hepatitis. Patients with chronic hepatitis C that have mixed cryoglobulinemia have an apparent duration of disease that is almost twice as long as those without cryoglobulinemia [89]. High prevalence of mixed cryoglobulinemia (35–90%) has been reported for patients with HCV infection [90,91]. However, in assessing the prevalence of mixed cryoglobulinemia it must be considered that prevalence has not been determined in populations of unselected HCV infected patients. Hence,

reports of high prevalence of mixed cryoglobulinemia may be due to a patient selection bias since most studies come from gastroenterology centers that have patient populations with an increased prevalence of patients with long duration disease, e.g. cirrhosis.

In only a minority of patients with mixed cryoglobulinemia are there extrahepatic clinical manifestations. In asymptomatic patients the significance, if any, of cryoglobulins in the development of extrahepatic clinical manifestations or cirrhosis has been the subject of speculation, particularly regarding cirrhosis. A recent meta-analysis of 19 studies on a total of 2323 patients with chronic hepatitis C found a 44% prevalence of cryoglobulinemia (analysis of the cryoglobulins was not provided) and a highly significant association between cirrhosis and cryoglobulinemia after adjustment for age, gender, and estimated duration of disease [91]. The authors concluded that cryoglobulins should be considered a prognostic indicator for the increased risk of cirrhosis in patients with chronic hepatitis C. This conclusion is obviously flawed since symptomatic type II cryoglobulinemia is associated with a low prevalence of cirrhosis [72]. Because of marked heterogeneity of cryoglobulins in HCV infection precise characterization of cryoglobulins and their specific role in the putative associated lesion must be elucidated to determine pathological significance. Thus far, there have been no studies to determine if there is a distinct type of cryoglobulin associated with cirrhosis. The role of specific cryoglobulins has been demonstrated, in part, only for the mRF in membranous proliferative glomerulonephritis lesions [92] and skin lesions [78] in HCV-type II cryoglobulinemia.

The hypothesis that with increasing duration of disease type III mixed cryoglobulinemia progresses to type II cryoglobulinemia was derived from the observation that the apparent duration of disease in patients with hepatitis C and type II cryoglobulins was twice as long as those with type III cryoglobulinemia [89]. More sensitive methods used for detection of monoclonal Ig [93] have revealed patients with oligoclonal IgM and has led to the additional postulation that the IgM RF producing B cells transform from polyclonality to oligoclonality to monoclonality with chronic infection [94]. There have been no studies to test this ‘cryoglobulin transition’ hypothesis.

The alternative interpretation of the data that underlies the cryoglobulin transition hypothesis is that polyclonal IgM RF, oligoclonal IgM, and monoclonal IgM RF (mRF) arise by distinct mechanisms and type II cryoglobulinemia develops without evolving from type III cryoglobulinemia through oligoclonal IgM RF populations of B cells. The mechanism for the production of mRF in HCV is not known. The majority of these mRF, WA mRF, are encoded by a restricted set of Ig genes that generated a unique antibody binding site epitope or crossidiotypic. MRF occur in other infections but the high prevalence of mRF and

presence of the WA crossidiotypic may be unique to HCV infection.

In HCV infection there is no molecular evidence that mRF B cells in patients with type II cryoglobulinemia arise from a population of polyclonal RF B cells in patients with type III cryoglobulinemia. On the contrary, recent studies using B cell clonal analysis in patients with chronic HCV infection have shown that the predominant mRF B cell in type II cryoglobulinemia, WA B cells, can be detected in approximately 4% of HCV-infected patients without cryoglobulinemia or RF but not in patients with type III cryoglobulinemia [95].

It is well established from animal model studies that circulating IgG immune complexes stimulates production of RF. The oligoclonal IgMs detected in HCV infection have not been shown to be RF. These oligoclonal IgM antibodies may represent a response to viral antigens rather than IgG immune complexes. Oligoclonal IgMs have been documented in other viral infections particularly in cerebrospinal fluid [96]. Moreover, there is no evidence that RF in human immune complex diseases transitions to mRF. In rheumatoid arthritis, the best example of an immune complex disease with RF formation, the development of mRF is an exceedingly rare occurrence.

### 3.2. Membranoproliferative glomerulonephritis (MPGN) independent of mixed cryoglobulinemia

The key issues regarding MPGN in patients with HCV infection were addressed at a recent NIH sponsored workshop on hepatitis C and renal disease [97]. These include reasons why some cryoglobulins are associated with kidney damage while others are not, and whether non-cryoglobulinemic associated glomerulonephritis can be caused by HCV. Glomerulonephritis, almost exclusively MPGN, is associated with mixed cryoglobulinemia secondary to HCV infection in approximately one third of patients [1]. The MPGN lesion is characterized by distinct morphological features: endocapillary proliferation; monocytic infiltration; double contour membranes; large eosinophilic; PAS-positive intraluminal deposits; and vasculitis of small and medium sized renal arteries [92]. On electron microscopy, deposits, usually subendothelial, are present and may have tubular and crystalline patterns that are also found in the cryoglobulins [98,99]. MPGN appears to occur predominantly with type II cryoglobulins [92,100].

The mRF in the type II cryoglobulins appears to be a critical element in the deposition of these cryoglobulins in glomeruli [92]. There is evidence from studies of MPGN associated with mixed cryoglobulins in patients with systemic lupus erythematosus that this may be a general role of mRF in MPGN lesions [101]. The relatively high prevalence of mRF in HCV infection may be the reason for the predominance of HCV in cryoglobulinemic glomerulonephritis [100].

Johnson and co-workers first proposed that MPGN occurs in HCV infected patients without cryoglobulinemia

[102]. However, in that initial study, five patients had cryoglobulinemia and all eight patients had RF in the serum. Moreover, in a second study by the Johnson group four patients that initially had MPGN without cryoglobulinemia developed cryoglobulinemia on follow-up [103]. This issue has become controversial since several studies have not found an association of HCV with MPGN [104–108] in the absence of cryoglobulinemia. Neither of the conflicting arguments is convincing [105,109] since definitive studies to detect and characterize RF in MPGN lesions in the absence of cryoglobulinemia, and B cell clonal analysis that can detect mRF B cells in the absence of cryoglobulinemia [71], have not been performed.

From studies in Italy [105], the United States [100,102,103], and Japan [107], MPGN associated with type II cryoglobulinemia is the predominant type of glomerulonephritis clinically associated with HCV infection. The prevalence of MPGN in HCV-type II cryoglobulinemia is approximately 30% [1]. However, from autopsy studies on 188 Japanese patients with HCV infection, predominantly with cirrhosis, the prevalence of histologic glomerular deposition of immune complexes was considerably higher than the prevalence of symptomatic glomerulonephritis [110]. The prevalence of histologic glomerulonephritis was 54.8%; the prevalence of MPGN, the most common type, was 11.2%. However, only 12.2% of patients, mainly with MPGN, had clinical manifestation of glomerulonephritis in the year prior to death. This finding is consistent with the higher prevalence of cryoglobulinemia among patients with advanced hepatitis C [89]. Considering the marked heterogeneity of immune complexes in HCV infection [111], a variety of immune complexes are most likely deposited in glomeruli, but only a portion of these, possibly those with mRF, appear to be nephrotoxic.

### 3.3. Infection of B cell and development of lymphomas

A widely held hypothesis is that the development of B cell malignancies in HCV-infected patients are related to infection of B cells by the virus. From the few studies reported on HCV in lymphoma cells, there is no definitive evidence that the virus is present in the malignant cells. In a study of eight HCV-infected patients with extranodal B-NHL, HCV was not detected in the lymphoma cells [112]. In a salivary gland lymphoma, the virus was found not in the lymphoma cells but in the non-malignant glandular epithelial cells [113]. In one of nine primary liver lymphomas in HCV-infected patients, HCV was detected in the lymphoma cells but not in surrounding hepatocytes using an in situ hybridization assay for HCV that had not been validated on known positive samples [114]. In a recent study on a patient with HCV infection, type II cryoglobulinemia and a monocytoid B cell lymphoma, evidence for replication of the virus, was demonstrated in the spleen and in a cell line developed



from the spleen. Since the cells were immortal in cell culture without employing hybridoma technology, the cell line was probably developed from the lymphoma cells. However identity of the cultured cells with lymphoma cells from the patient was not demonstrated [115].

In opposition to a direct role of HCV in lymphomagenesis is that only 4–6% of patients with HCV-type II cryoglobulinemia develop frank B cell malignancies in long-term follow-up studies [116,117]. Broader studies on the association of B-NHL with HCV infection are conflicting. Eight studies, performed mainly in Italy and Japan, found increased prevalence of HCV infection in patients with B-NHL (9–32%) compared to controls (0–3.1%) [118–123] but ten studies done mainly in the United Kingdom and North America did not find an association [124–131]. However in a longitudinal study of 2162 chronic hepatitis patients in Osaka between 1957 and 1997, the region having the highest prevalence of HCV infection in Japan, only four patients with B-NHL were detected [122]. Hence, HCV infection may be associated with B-NHL but not as prominently as suggested by some of the studies from Italy and Japan. The very high prevalence detected in the latter studies may be due to confounding patient selection bias because of the endemic regions in Italy and Japan. On the other hand, the low prevalence in the non-endemic areas may be due to inadequate patient sample sizes considering the low HCV prevalence in these regions.

A role for HCV in lymphomagenesis is not readily apparent since HCV is not an oncogenic virus. However, such a role cannot be excluded since the HCV core protein has been reported to be involved in transformation of cells to malignant phenotypes [132]. In addition, hypermutation of immunoglobulin and oncogenes in HCV-infected cells has been demonstrated in a preliminary report [133]. The recent finding that regression of splenic lymphoma associated with HCV infection occurs synchronously with decline in viremia with anti-viral therapy strongly suggests a role for HCV in the development of malignancy [134]. However, thus far, no specific HCV lymphomagenic mechanisms have been delineated.

The evidence on HCV lymphotropism is conflicting [59,135]. It has not been established that HCV has an unrestricted tropism for lymphocytes. The data supporting lymphotropism is extensively reviewed in reference [59]. The sum of the evidence suggests that replication of HCV occurs only in a small number of peripheral blood mononuclear cells (PBMC), and probably only under abnormal conditions, i.e. immunosuppression [136,137], cell transformation [115]. The finding, in several studies, that the quasispecies of HCV present in PBMC were not present in the serum is consistent with this assessment [137–139]. If HCV infection of lymphocytes was as productive as that of hepatocytes then the virus produced by these cells would constitute a significant part of the viremia since the blood lymphocyte compartment is  $1 \times 10^{10}$  and lymphocyte traffic through the blood is

$5 \times 10^{11}$  cells. The latter number equals the number of lymphocytes contained within the whole human body and is 2.5 times the number of hepatocytes in the liver [140].

Chromosomal translocation has been suggested as a possible mechanism for the role of HCV infection in lymphomagenesis. A high prevalence of  $t(14;18)$  translocation in patients with HCV infection and an even higher prevalence in patients with HCV-type II cryoglobulinemia have been reported [141–143]. This translocation is thought to predispose to malignancy because the Bcl-2 over-expression that may accompany the translocation confers longevity on cells. However, thus far,  $t(14;18)$  translocation and Bcl-2 over expression in HCV infected patients have not been demonstrated to occur in the same cells and more specifically in the mRF producing B cells [141,144]. This is a critical point since Bcl-2 over expression may be a characteristic of monoclonal B cells in patients with HCV-type II cryoglobulinemia [145] and thus far, all WA mRF cells studied have shown Bcl-2 over-expression without  $t(14;18)$  translocation [146,147]. Since Bcl-2 over expression does not always accompany  $t(14;18)$  translocation [148], B cells with  $t(14;18)$  translocation without Bcl-2 over-expression and mRF B cells that overexpress Bcl-2 without  $t(14;18)$  may present in the blood. The two populations could not be discerned without isolation studies that thus far, have not been performed.

A review of 854 lymphomas reported in patients with HCV infection (Table 2) [37,38,43,47,112,118–121,126,128,149–182] illustrates the predominance of B-NHL in these patients. Seventy-nine per cent of patients tested for cryoglobulinemia (42% were not tested) were positive suggesting a higher prevalence of B-NHL among patients with than without cryoglobulinemia. In a multicenter case-control study included in Table 2, the HCV-attributable risk for developing B-NHL was estimated at 4.6%, i.e. approximately one in twenty B-NHL in Italy are associated with HCV infection [183].

The  $t(14;18)$  translocation, first discovered in patients with follicular cell lymphoma, is present in most patients with this lymphoma. However,  $t(14;18)$  translocation may occur in high prevalence in normal individuals, increase in prevalence with aging, and occur with greater frequency in proliferating cells [184–188]. Hence,  $t(14;18)$  translocation in HCV infection does not necessarily have implications for lymphomagenesis. If this transformation does have a role, an increase percentage of B-NHL would be expected to be follicular lymphoma especially among patients with HCV-type II cryoglobulinemia. Table 3 addresses this question. The percentage of each subtype of B-NHL in the various groups of HCV-infected patients from 45% of the patients in Table 2 (55% of B-NHL were not subtyped) are compared to distribution of these subtypes in the general population obtained from an international lymphoma study [189]. Contrary to expectation, immunoplasmaeytic lymphomas/Waldenstrom macroglobulinemia and not follicular lymphoma appears to be the most common B-NHL among

**Table 2**  
**Predominance of B-NHL lymphomas in HCV-infected patients with and without cryoglobulinemia [37,38,43,47,112,118–121,126,128,149–182]**

Lymphoma type	Symptomatic type II cryo	Type II cryo	Type III cryo	Cryo not typed	Cryo negative	Cryo not tested
Lymphoplasmacytic/Waldenstrom	12	10	0	23	4	2
Chronic lymphocytic/small cell	3	0	0	0	0	0
Marginal zone cell						
Nodal	2	2	3	11	3	13
Extranodal	2	0	0	6	36	34
Splenic	0	0	0	7	2	0
Follicular	2	0	0	14	17	5
Mantle cell	0	1	1	0	1	0
Diffuse large B cell	7	4	7	16	22	32
Plasma cell myeloma	0	0	0	0	0	5
Burkitt's	0	0	0	0	1	0
Hairy cell leukemia	0	0	0	2	0	0
Hodgkin's	0	0	0	0	1	1
NK	0	0	0	0	0	0
T-cell	0	0	0	2	1	0
B-NHL, unclassified	19	0	0	267	44	158
Unclassifiable	0	0	0	0	0	3

Classification, ref. [149].

patients with type II cryoglobulinemia. Follicular lymphoma that constitutes 22% of all B-NHL was not increased in any of the HCV infected groups, but rather there was a significant decreased percentage in the groups with cryoglobulinemia. Also of note, there was an increased percentage of extra-nodal marginal zone cell lymphomas, in particular, among HCV-infected patients without cryoglobulins.

The increased percentage of extra-nodal marginal cell lymphomas in HCV infection illustrated in Table 3 and the report of more than a 10-fold greater risk for developing B-NHL of the liver and salivary gland than the risk of lymphomas at other sites is consistent with the hypothesis that B-NHL arise selectively from marginal zone B cell

[190]. The extra-nodal marginal zone cell lymphomas are considered to be derived from organized lymphoid tissue that accumulate in response to infection or as a component of autoimmune disease at a site beyond the lymph node [191]. There is evidence that the monoclonal B cells in type II cryoglobulinemia arise predominantly in the pseudo-follicle in the liver [145,192]. Also consistent with this hypothesis are studies on a patient with partial trisomy 3 and HCV-type II cryoglobulinemia. Phenotypic markers characteristic of marginal zone cells were present on the WA B cells present in the blood in this patient [147]. The patient had features of splenic lymphoma and, like the monoclonal B cells patients with splenic lymphoma and HCV-infection, the WA B cell regressed with antiviral

**Table 3**  
**Percentage of B-NHL subtypes among patient in Table 2: comparison with International Lymphoma Study [189]**

Lymphoma type	International Lymphoma Study	Table 2		
		Sym type II	All cryo	Neg cryo
Lymphoplasmacytic/Waldenstrom	1.2	43*****	33***	5
Chronic lymphocytic/small cell	6.7	11	1*	0**
Marginal zone cell				
Nodal	1.8	7	13*****	3
Extranodal	7.6	7	9	42*****
Splenic	<1	0	5***	2
Follicular	22.1	7*	12****	20
Mantle cell	6.0	0	1*	1*
Diffuse large B cell	30.6	25	25**	26*
Burkitt's	<1	0	0	0
Total patients	1069	28	137	85

Two proportion analysis (PASS 2000 software):  $P = * < 0.05$ ,  $** < 0.01$ ,  $*** < 0.001$ ,  $**** < 0.0001$ ,  $***** < 10^{-6}$ .

therapy. The over-represented region of chromosome 3, bands 3q11–29, that were present in this patient is commonly involved in marginal zone B cell lymphomas. The prevalence of this chromosomal transformation in HCV infection has not been determined.

Approximately 25% of B-NHLs in patients with HCV-type II cryoglobulinemia appeared to arise from WA B cells [150,193]. Based on the reactivity of HCV envelope E2 protein with CD81 and homologies of Ig V genes sequences of lymphoma cells with mRF and anti-E2 sequences, it has been postulated that E2 drives proliferation of the monoclonal B cells in patients with HCV-type II cryoglobulinemia that undergo malignant transformation to B-NHL [150,194]. However, the rationale for this hypothesis is faulty since none of the WA mRF sequences, the major mRF in HCV-type II cryoglobulinemia, have any significant homology with the cloned anti-E2 envelope antibodies [193] and the expressed receptors of the with IgG [195]. Moreover, it has been demonstrated that WA mRF does not cross-react with E2 (Elfahal M, Agnello V, unpublished observations).

At present there are more questions than answers regarding the association of B-NHL with HCV infection. The critical question is whether virus replication occurs in normal B cells and is directly lymphomagenic or is lymphomagenesis a stochastic process accompanying HCV-driven proliferation of B cells. Unequivocal evidence of replication of HCV in transformed B cells has been presented [115], however, there is no unequivocal evidence that replication occurs in normal B cells. The low prevalence of B-NHL in long term follow-up studies on patients with HCV-type II cryoglobulinemia favors the stochastic hypothesis [116,117].

## Acknowledgements

This study was supported in part by a Veterans Affairs Department Merit Review Grant. Dr De Rosa was a fellow of the Robert E. Wise M.D. Research and Education Institute. We thank Carol Spencer for editing the manuscript.

## References

- [1] Agnello V. Mixed cryoglobulinemia and other extrahepatic manifestations of HCV infection. In: Liang TJ, Hoofnagle JH, editors. Hepatitis C. New York: Academic Press; 2000. p. 295–315.
- [2] Munoz-Rodriguez FJ, Tassies D, Font J, Reverter JC, Cervera R, Sanchez-Tapias JM, et al. Prevalence of hepatitis C virus infection in patients with antiphospholipid syndrome. *J Hepatol* 1999;30:770–773.
- [3] Prieto J, Yuste JR, Beloqui O, Civeira MP, Riezu JI, Aguirre B, et al. Anticardiolipin antibodies in chronic hepatitis C: implication of hepatitis C virus as the cause of the antiphospholipid syndrome. *Hepatology* 1996;23:199–204.
- [4] Gruber A, Grillner L, Norder H, Magnus L, Bjorkholm M. Severe aplastic anemia associated with seronegative community-acquired hepatitis C virus infection. *Ann Hematol* 1993;66:157–159.
- [5] Hibbs JR, Issaragrisil S, Young NS. High prevalence of hepatitis C viremia among aplastic anemia patients and controls from Thailand. *Am J Trop Med Hyg* 1992;46:564–570.
- [6] Moccia F, Tognoni E, Boccaccio P. Autoimmune hemolytic anemia in chronic hepatitis C virus infection: an unusual extrahepatic autoimmune manifestation. *Ann Ital Med Int* 2001;16:256–259.
- [7] Srinivasan R. Autoimmune hemolytic anemia in treatment-naive chronic hepatitis C infection. *J Clin Gastroenterol* 2001;32:245–247.
- [8] Tran A, Quaranta JF, Benzaken S, Thiers V, Chau HT, Hastier P, et al. High prevalence of thyroid autoantibodies in a prospective series of patients with chronic hepatitis C before interferon therapy. *Hepatology* 1993;18:253–257.
- [9] Ganne-Carrie N, Medini A, Coderc E, Seror O, Christidis C, Grimbirt S, et al. Latent autoimmune thyroiditis in untreated patients with HCV chronic hepatitis: a case-control study. *J Autoimmun* 2000;14:189–193.
- [10] Rocco A, Gargano S, Provenzano A, Nardone M, De Sanctis GM, Altavilla N, et al. Incidence of autoimmune thyroiditis in interferon-alpha treated and untreated patients with chronic hepatitis C virus infection. *Neuroendocrinol Lett* 2001;22:39–44.
- [11] Forton DM, Allsop JM, Main J, Foster GR, Thomas HC, Taylor-Robinson SD. Evidence for a cerebral effect of the hepatitis C virus. *Lancet* 2001;358:38–39.
- [12] Kramer L, Bauer E, Funk G, Hofer H, Jessner W, Steindl-Munda P, et al. Subclinical impairment of brain function in chronic hepatitis C infection. *J Hepatol* 2002;37:349–354.
- [13] Gordon SC. Extrahepatic manifestations of hepatitis C. *Dig Dis* 1996;14:157–168.
- [14] Ilter N, Senol E, Gurer MA, Oztas MO. Behcet's disease and HCV infection. *Int J Dermatol* 2000;39:396–397.
- [15] Munke H, Stockmann F, Ramadori G. Possible association between Behcet's syndrome and chronic hepatitis C virus infection. *N Engl J Med* 1995;332:400–401.
- [16] Ishizaka Y, Ishizaka N, Takahashi E, Tooda E, Hashimoto H, Nagai R, et al. Association between hepatitis C virus seropositivity, carotid-artery plaque, and intima-media thickening. *Lancet* 2002;359:133–135.
- [17] Ishizaka Y, Ishizaka N, Takahashi E, Unuma T, Tooda E, Hashimoto H, et al. Association between hepatitis C virus core protein and carotid atherosclerosis. *Circ J* 2003;67:26–30.
- [18] Pawlotsky JM, Ben Yahia M, Andre C, Voisin MC, Intrator L, Roudot-Thoraval F, et al. Immunological disorders in C virus chronic active hepatitis: a prospective case-control study. *Hepatology* 1994;19:841–848.
- [19] Crowson AN, Nuovo G, Ferri C, Magro CM. The dermatopathologic manifestations of hepatitis C infection: a clinical, histological, and molecular assessment of 35 cases. *Hum Pathol* 2003;34:573–579.
- [20] Nishikai M, Miyairi M, Kosaka S. Dermatomyositis following infection with hepatitis C virus. *J Rheumatol* 1994;21:1584–1585.
- [21] Arai M, Murase K, Kusakabe A, Yoshioka K, Fukuzawa Y, Ishikawa T, et al. Prevalence of diabetes mellitus in Japanese patients infected chronically with hepatitis C virus. *J Gastroenterol* 2003;38:355–360.
- [22] Mehta SH, Strathdee SA, Thomas DL. Association between hepatitis C virus infection and diabetes mellitus. *Epidemiol Rev* 2001;23:302–312.
- [23] Buskila D, Shnaider A, Neumann L, Lorber M, Zilberman D, Hilzenrat N, et al. Musculoskeletal manifestations and autoantibody profile in 90 hepatitis C virus infected Israeli patients. *Semin Arthritis Rheum* 1998;28:107–113.
- [24] Buskila D, Shnaider A, Neumann L, Zilberman D, Hilzenrat N, Sikuler E. Fibromyalgia in hepatitis C virus infection. Another infectious disease relationship. *Arch Intern Med* 1997;157:2497–2500.
- [25] Lacaille F, Zylberberg H, Hagege H, Roualdes B, Meyrignac C, Chousterman M, et al. Hepatitis C associated with Guillain-Barre syndrome. *Liver* 1998;18:49–51.

- [26] Matsumori A, Ohashi N, Nishio R, Kakio T, Hara M, Furukawa Y, et al. Apical hypertrophic cardiomyopathy and hepatitis C virus infection. *Jpn Circ J* 1999;63:433–438.
- [27] Matsumori A, Ohashi N, Hasegawa K, Sasayama S, Eto T, Imaizumi T, et al. Hepatitis C virus infection and heart diseases: a multicenter study in Japan. *Jpn Circ J* 1998;62:389–391.
- [28] Matsumori A, Matoba Y, Nishio R, Shioi T, Ono K, Sasayama S. Detection of hepatitis C virus RNA from the heart of patients with hypertrophic cardiomyopathy. *Biochem Biophys Res Commun* 1996;222:678–682.
- [29] Petit JM, Benichou M, Duvillard L, Jooste V, Bour JB, Minello A, et al. Hepatitis C virus-associated hypobetalipoproteinemia is correlated with plasma viral load, steatosis, and liver fibrosis. *Am J Gastroenterol* 2003;98:1150–1154.
- [30] Ferri C, La Civita L, Fazzi P, Solfanelli S, Lombardini F, Begliomini E, et al. Interstitial lung fibrosis and rheumatic disorders in patients with hepatitis C virus infection. *Br J Rheumatol* 1997;36:360–365.
- [31] Arase Y, Ikeda K, Tsubota A, Suzuki Y, Saitoh S, Kobayashi M, et al. Serum levels of gamma-globulin and total bilirubin influence the prevalence of multiple extrahepatic complication in patients with hepatitis C virus infection. *Hepatology* 2003;37:14–21.
- [32] Pockros PJ, Duchini A, McMillan R, Nyberg LM, McHutchison J, Vierns E. Immune thrombocytopenic purpura in patients with chronic hepatitis C virus infection. *Am J Gastroenterol* 2002;97:2040–2045.
- [33] Sakuraya M, Murakami H, Uchiumi H, Hatsumi N, Akiba T, Yokohama A, et al. Steroid-refractory chronic idiopathic thrombocytopenic purpura associated with hepatitis C virus infection. *Eur J Haematol* 2002;68:49–53.
- [34] Ilan Y, Shouval D, Ashur Y, Manns M, Naparstek Y. IgA deficiency associated with chronic hepatitis C virus infection. A cause or an effect? *Arch Intern Med* 1993;153:1588–1592.
- [35] Figueiredo LC, Carrilho FJ, de Andrade HF, Migliari DA. Oral lichen planus and hepatitis C virus infection. *Oral Dis* 2002;8:42–46.
- [36] Imhof M, Popal H, Lee JH, Zeuzem S, Milbradt R. Prevalence of hepatitis C virus antibodies and evaluation of hepatitis C virus genotypes in patients with lichen planus. *Dermatology* 1997;195:1–5.
- [37] Tursi A, Brandimante G, Chiarelli F, Spagnoli A, Torello M. Detection of HCV RNA in gastric mucosa-associated lymphoid tissue by in situ hybridization: evidence of a new extrahepatic localization of HCV with increased risk of gastric malt lymphoma. *Am J Gastroenterol* 2002;97:1802–1806.
- [38] Luppi M, Longo G, Ferrari MG, Ferrara L, Marasca R, Barozzi P, et al. Additional neoplasms and HCV infection in low-grade lymphoma of MALT type. *Br J Haematol* 1996;94:373–375.
- [39] Bandi L. Renal manifestations of hepatitis C virus infection. Extrahepatic complications often are silent – and thus overlooked. *Postgrad Med* 2003;113:73–76, 86.
- [40] Hoch B, Juknevičius I, Liapis H. Glomerular injury associated with hepatitis C infection: a correlation with blood and tissue HCV-PCR. *Semin Diagn Pathol* 2002;19:175–187.
- [41] Johnson RJ, Gretch DR, Yamabe H, Hart J, Bacchi CE, Hartwell P, et al. Membranoproliferative glomerulonephritis associated with hepatitis C virus infection. *N Engl J Med* 1993;328:465–470.
- [42] Rollino C, Roccatello D, Giachino O, Basolo B, Piccoli G. Hepatitis C virus infection and membranous glomerulonephritis. *Nephron* 1991;59:319–320.
- [43] Agnello V, Chung RT, Kaplan LM. A role for hepatitis C virus infection in type II mixed cryoglobulinemia. *N Engl J Med* 1992;327:1490–1495.
- [44] Ferri C, La Civita L, Longombardo G, Greco F, Bombardieri S. Hepatitis C virus and mixed cryoglobulinaemia. *Eur J Clin Invest* 1993;23:399–405.
- [45] Wilson SE, Lee WM, Murakami C, Weng J, Moninger GA. Mooren-type hepatitis C virus-associated corneal ulceration. *Ophthalmology* 1994;101:736–745.
- [46] Montella M, Crispo A, Russo F, Ronga D, Tridente V, Tamburini M. Hepatitis C virus infection and new association with extrahepatic disease: multiple myeloma. *Haematologica* 2000;85:883–884.
- [47] Gharagozloo S, Khoshnoodi J, Shokri F. Hepatitis C virus infection in patients with essential mixed cryoglobulinemia, multiple myeloma and chronic lymphocytic leukemia. *Pathol Oncol Res* 2001;7:135–139.
- [48] Iannitto E, Barbera V, Gambino R, Ammatuna E, Montalto G, Di Stefano R. Higher frequency of HCV in patients with non-Hodgkin lymphoma: is it enough to suggest an association with B-cell NHL? *Hepatology* 2003;37:481–482.
- [49] Chindamo MC, Spector N, Segadas JA, Pimenta G, Vanderborcht B, Morais JC, et al. Prevalence of hepatitis C infection in patients with non-Hodgkin's lymphomas. *Oncol Rep* 2002;9:657–659.
- [50] Alvares-Da-Silva MR, Francisoni CF, Waechter FL. Acute hepatitis C complicated by pancreatitis: another extrahepatic manifestation of hepatitis C virus? *J Viral Hepat* 2000;7:84–86.
- [51] Carson CW, Conn DL, Czaja AJ, Wright TL, Brecher ME. Frequency and significance of antibodies to hepatitis C virus in polyarteritis nodosa. *J Rheumatol* 1993;20:304–309.
- [52] Quint L, Deny P, Guillemin L, Granger B, Jarrousse B, Lhote F, et al. Hepatitis C virus in patients with polyarteritis nodosa. Prevalence in 38 patients. *Clin Exp Rheumatol* 1991;9:253–257.
- [53] Aisa Y, Yokomori H, Kashiwagi K, Nagata S, Yanagisawa R, Takahashi M, et al. Polymyositis, pulmonary fibrosis and malignant lymphoma associated with hepatitis C virus infection. *Intern Med* 2001;40:1109–1112.
- [54] Weidensaul D, Imam T, Holyst MM, King PD, McMurray RW. Polymyositis, pulmonary fibrosis, and hepatitis C. *Arthritis Rheum* 1995 Mar;38:437–439.
- [55] Navas S, Bosch O, Castillo I, Marriott E, Carreno V. Porphyria cutanea tarda and hepatitis C and B viruses infection: a retrospective study. *Hepatology* 1995;21:279–284.
- [56] DeCastro M, Sanchez J, Herrera JF, Chaves A, Duran R, Garcia-Buey L, et al. Hepatitis C virus antibodies and liver disease in patients with porphyria cutanea tarda. *Hepatology* 1993;17:551–557.
- [57] Fargion S, Piperno A, Cappellini MD, Sampietro M, Fracanzani AL, Romano R, et al. Hepatitis C virus and porphyria cutanea tarda: evidence of a strong association. *Hepatology* 1992;16:1322–1326.
- [58] Hsu FC, Starkebaum G, Boyko EJ, Dominitz JA. Prevalence of rheumatoid arthritis and hepatitis C in those age 60 and older in a US population based study. *J Rheumatol* 2003;30:455–458.
- [59] Zignego AL, Brechot C. Extrahepatic manifestations of HCV infection: facts and controversies. *J Hepatol* 1999;31:369–376.
- [60] Carrozzo M, Gandolfo S. Oral diseases possibly associated with hepatitis C virus. *Crit Rev Oral Biol Med* 2003;14:115–127.
- [61] Pirisi M, Scott C, Fabris C, Ferraccioli G, Soardo G, Ricci R, et al. Mild sialoadenitis: a common finding in patients with hepatitis C virus infection. *Scand J Gastroenterol* 1994;29:940–942.
- [62] Nagao Y, Hanada S, Shishido S, Ide T, Kumashiro R, Ueno T, et al. Incidence of Sjögren's syndrome in Japanese patients with hepatitis C virus infection. *J Gastroenterol Hepatol* 2003;18:258–266.
- [63] Mariette X, Zerbib M, Jaccard A, Schenmetzler C, Danon F, Clauvel JP. Hepatitis C virus and Sjögren's syndrome. *Arthritis Rheum* 1993;36:280–281.
- [64] Ramos-Casals M, Font J, Garcia-Carrasco M, Cervera R, Jimenez S, Trejo O, et al. Hepatitis C virus infection mimicking systemic lupus erythematosus: study of hepatitis C virus infection in a series of 134 Spanish patients with systemic lupus erythematosus. *Arthritis Rheum* 2000;43:2801–2806.
- [65] McMurray RW. Hepatitis C-associated autoimmune disorders. *Rheum Dis Clin North Am* 1998;24:353–374.
- [66] Hadziyannis SJ. Non-hepatic manifestations and combined diseases in HCV infection. *Dig Dis Sci* 1996;41:635–745.
- [67] Gordon SC. Extrahepatic manifestations of hepatitis C. *Dig Dis* 1996;14:157–168.



- [68] Izumi T, Sasaki R, Tsunoda S, Akutsu M, Okamoto H, Miura Y. B cell malignancy and hepatitis C virus infection. *Leukemia* 1997;11: 516–518.
- [69] Izumi T, Sasaki R, Shimizu R, Miyazato A, Hoshino Y, Miura Y, et al. Hepatitis C virus infection in Waldenström's macroglobulinemia. *Am J Hematol* 1996;52:238–239.
- [70] Strassburg CP, Obermayer-Straub P, Manns MP. Autoimmunity in hepatitis C and D virus infection. *J Viral Hepat* 1996;3:49–59.
- [71] Gao L, Knight GB, Agnello V. Molecular analysis of clonal B cell expansion in HCV infection: a potential marker for development of type II mixed cryoglobulinemia (abstr). *Antiviral Ther* 2000;5:C42.
- [72] Monti G, Galli M, Invernizzi F, Pioltelli P, Saccardo F, Monteverde A. Cryoglobulinaemias: a multi-centre study of the early clinical and laboratory manifestations of primary and secondary disease. GISC. Italian Group for the Study of Cryoglobulinaemias. *QJM* 1995;88: 115–126.
- [73] Haddad J, Deny P, Munz-Gotheil C, Ambrosini JC, Trinchet JC, Pateron D, et al. Lymphocytic sialadenitis of Sjögren's syndrome associated with chronic hepatitis C virus liver disease. *Lancet* 1992; 339:321–323.
- [74] Roy K, Bagg J. Hepatitis C virus and oral disease: a critical review. *Oral Dis* 1999;5:270–277.
- [75] King PD, McMurray RW, Becherer PR. Sjögren's syndrome without mixed cryoglobulinemia is not associated with hepatitis C virus infection. *Am J Gastroenterol* 1994;89:1047–1050.
- [76] Arrieta JJ, Rodriguez-Inigo E, Ortiz-Movilla N, Bartolome J, Pardo M, Manzarbeitia F, et al. In situ detection of hepatitis C virus RNA in salivary glands. *Am J Pathol* 2001;158:259–264.
- [77] Taliani G, Celestino D, Badolato MC, Pennica A, Bozza A, Poliandri G, et al. Hepatitis C virus infection of salivary gland epithelial cells. Lack of evidence. *J Hepatol* 1997;26:1200–1206.
- [78] Agnello V, Abel G. Localization of hepatitis C virus in cutaneous vasculitic lesions in patients with type II cryoglobulinemia. *Arthritis Rheum* 1997;40:2007–2015.
- [79] Agnello V, Abel G, Elfahal M, Knight GB, Zhang QX. Hepatitis C virus and other flaviviridae viruses enter cells via low density lipoprotein receptor. *Proc Natl Acad Sci USA* 1999;96:12766–12771.
- [80] Kioke K, Moriya K, Ishibashi K, Yotsuyanagi H, Shintani Y, Fujie H, et al. Sialadenitis histologically resembling Sjögren syndrome in mice transgenic for hepatitis C virus envelope genes. *Proc Natl Acad Sci USA* 1997;94:233–236.
- [81] Agnello V, Liang TJ. Is hepatitis C a sialodacryoadentitis virus? *Hepatology* 1997;26:509–510.
- [82] Mignogna MD, Fedele S, Lo Russo L, Ruoppo E, Adamo D, Lo Muzio L. Extrahepatic manifestations of hepatitis C virus infection: the slowly unraveling picture of oral lichen planus. *J Hepatol* 2002; 37:412–413.
- [83] Pilli M, Penna A, Zerbinì A, Vescovi P, Manfredi M, Negro F, et al. Oral lichen planus pathogenesis: a role for the HCV-specific cellular immune response. *Hepatology* 2002;36:1446–1452.
- [84] Bloomer JR. Chronic hepatitis C and porphyria cutanea tarda. In: Liang TJ, Hoofnagle JH, editors. *Hepatitis C*. New York: Academic Press; 2000. p. 351–361.
- [85] Bonkovsky HL, Poh-Fitzpatrick M, Pimstone N, Obando J, DiBisceglie A, Tattrie C, et al. Porphyria cutanea tarda, hepatitis C, and HFE gene mutations in North America. *Hepatology* 1998;27: 1661–1669.
- [86] Moran MJ, Fontanellas A, Brudieux E, Hombrados I, de Ledinghen V, Couzigou P, et al. Hepatic uroporphyrinogen decarboxylase activity in porphyria cutanea tarda patients: the influence of virus C infection. *Hepatology* 1998;27:584–589.
- [87] Meltzer M, Franklin EC, Elias K, McCluskey RT, Cooper N. Cryoglobulinemia – a clinical and laboratory study. II. Cryoglobulins with rheumatoid factor activity. *Am J Med* 1966;40:837–856.
- [88] Brouet JC, Clauvel JP, Danon F, Klein M, Seligmann M. Biologic and clinical significance of cryoglobulins. A report of 86 cases. *Am J Med* 1974;57:775–788.
- [89] Lunel F, Musset L, Cacoub P, Frangeul L, Cresta P, Perrin M, et al. Cryoglobulinemia in chronic liver diseases: role of hepatitis C virus and liver damage. *Gastroenterology* 1994;106:1291–1300.
- [90] Lunel F, Musset L. Mixed cryoglobulinemia and hepatitis C virus infection. *Minerva Med* 2001;92:35–42.
- [91] Kayali Z, Buckwold VE, Zimmerman B, Schmidt WN. Hepatitis C, cryoglobulinemia, and cirrhosis: a meta-analysis. *Hepatology* 2002; 36:978–985.
- [92] Fornasieri A, D'Amico G. Type II mixed cryoglobulinemia, hepatitis C infection, and glomerulonephritis. *Nephrol Dial Transplant* 1996; 11:25–30.
- [93] Musset L, Diemert MC, Taibi F, Thi Huong Du L, Cacoub P, Leger JM, et al. Characterization of cryoglobulins by immunoblotting. *Clin Chem* 1992;38:798–802.
- [94] Schifferli JA, French LE, Tissot JD. Hepatitis C virus infection, cryoglobulinemia, and glomerulonephritis. *Adv Nephrol Necker Hosp* 1995;24:107–129.
- [95] Gao L, Knight GB, Agnello V. Molecular analysis of clonal B cell expansion in HCV infection: a potential marker for development of type-2 mixed cryoglobulinemia (abstr). *Antiviral Ther* 2000;5:C42.
- [96] Franciotta D, Zardini E, Bono G, Brustia R, Minoli L, Cosi V. Antigen-specific oligoclonal IgG in AIDS-related cytomegalovirus and toxoplasma encephalitis. *Acta Neurol Scand* 1996;94:215–218.
- [97] Meyers CM, Seeff LB, Stehman-Breen CO, Hoofnagle JH. Hepatitis C and renal disease: an update. *Am J Kidney Dis* 2003; 42:631–657.
- [98] Cordonnier D, Martin H, Gros Lambert P, Micouin C, Chenais F, Stoebner P. Mixed IgG-IgM cryoglobulinemia with glomerulonephritis. Immunochemical, fluorescent and ultrastructural study of kidney and in vitro cryoprecipitate. *Am J Med* 1975;59:867–872.
- [99] Stoebner P, Renversez JC, Groude J, Vialtel P, Cordonnier D. Ultrastructural study of human IgG and IgG-IgM crystalcryoglobulins. *Am J Clin Pathol* 1979;71:404–410.
- [100] Pucillo LP, Agnello V. Membranoproliferative glomerulonephritis associated with hepatitis B and C viral infections: from virus like particles in the cryoprecipitate to viral localization in paramesangial deposits, problematic investigations prone to artifacts. *Curr Opin Nephrol Hypertens* 1994;3:465–470.
- [101] Agnello V, Koffler D, Kunkel HG. Immune complex systems in the nephritis of systemic lupus erythematosus. *Kidney Int* 1973;3:90–99.
- [102] Johnson RJ, Gretch DR, Yamabe H, Hart J, Bacchi CE, Hartwell P, et al. Membranoproliferative glomerulonephritis associated with hepatitis C virus infection. *N Engl J Med* 1993;328:465–470.
- [103] Johnson RJ, Gretch DR, Couser WG, Alpers CE, Wilson J, Chung M, et al. Hepatitis C virus-associated glomerulonephritis. Effect of  $\alpha$ -interferon therapy. *Kidney Int* 1994;46:1700–1774.
- [104] Rostoker G, Deforges L, Ben Maadi A, Remy P, Bourgeon B, Lang P, et al. Low prevalence of hepatitis C virus antibodies among adult patients with primary glomerulonephritis in France. *Nephron* 1993; 63:367.
- [105] D'Amico G, Fornasieri A. Cryoglobulinemic glomerulonephritis: a membranoproliferative glomerulonephritis induced by hepatitis C virus. *Am J Kidney Dis* 1995;25:361–369.
- [106] Lai FM, Tam JS, Liew CT, Ip M, Lai KN. Low prevalence of hepatitis C virus antibodies with primary membranous nephropathy and membranoproliferative glomerulonephritis in Hong Kong. *Nephron* 1995;70:367–368.
- [107] Yamabe H, Johnson RJ, Gretch DR, Fukushi K, Osawa H, Miyata M, et al. Hepatitis C virus infection and membranoproliferative glomerulonephritis in Japan. *J Am Soc Nephrol* 1995;6:220–223.
- [108] Cosio FG, Roche Z, Agarwal A, Falkenhain ME, Sedmak DD, Fergusson RM. Prevalence of hepatitis C in patients with idiopathic glomerulopathies in native and transplant kidneys. *Am J Kidney Dis* 1996;28:752–758.
- [109] Stehman-Breen C, Alpers CE, Willson R, Johnson RJ. Is there a hepatitis C virus-associated membranoproliferative glomerulonephritis? *Am J of Kidney Dis* 1997;30:589–590.

- [110] Arase Y, Ikeda K, Murashima N, Chayama K, Tsubota A, Koida I, et al. Glomerulonephritis in autopsy cases with hepatitis C virus infection. *Intern Med* 1998;37:836–840.
- [111] Agnello V. Mixed cryoglobulinaemia after hepatitis C virus: more and less ambiguity. *Ann Rheum Dis* 1998;57:701–702.
- [112] Ascoli V, Lo Coco F, Artini M, Levrero M, Martelli M, Negro F. Extranodal lymphomas associated with hepatitis C virus infection. *Am J Clin Pathol* 1998;109:600–609.
- [113] De Vita S, Sansonno D, Dolcetti R, Ferraccioli G, Carbone A, Cornacchiulo V, et al. Hepatitis C virus within a malignant lymphoma lesion in the course of type II mixed cryoglobulinemia. *Blood* 1892;86:1887–1887.
- [114] Ohsawa M, Tomita Y, Hashimoto M, Kanno H, Aozasa K. Hepatitis C viral genome in a subset of primary hepatic lymphomas. *Mod Pathol* 1998;11:471–478.
- [115] Sung VM, Shimodaira S, Doughty AL, Picchio GR, Can H, Yen TS, et al. Establishment of B-cell lymphoma cell lines persistently infected with hepatitis C virus in vivo and in vitro: the apoptotic effects of virus infection. *J Virol* 2003;77:2134–2146.
- [116] Gorevic PD, Frangione B. Mixed cryoglobulinemia cross-reactive idiotypes: implications for the relationship of MC to rheumatic and lymphoproliferative diseases. *Semin Hematol* 1991;28:79–94.
- [117] Invernizzi F, Galli M, Serino G, Monti G, Meroni LP, Granatieri C. Secondary and essential cryoglobulinemias. Frequency, nosological classification, and long-term follow-up. *Acta Haematol* 1983;70:73–82.
- [118] Silvestri F, Pipan C, Barillari G, Zaja F, Fanin R, Infanti L, et al. Prevalence of hepatitis C virus infection in patients with lymphoproliferative disorders. *Blood* 1996;87:4296–4301.
- [119] Kuniyoshi M, Nakamura M, Sakai H, Enjoji M, Kinukawa N, Kotoh K, et al. Prevalence of hepatitis B or C virus infections in patients with non-Hodgkin's lymphoma. *J Gastroenterol Hepatol* 2001;16:215–219.
- [120] Zuckerman E, Zuckerman T, Levine AM, Douer D, Gutekunst K, Mizokami M, et al. Hepatitis C virus infection in patients with B-cell non-Hodgkin lymphoma. *Ann Intern Med* 1997;127:423–428.
- [121] Luppi M, Longo G, Ferrari MG, Barozzi P, Marasca R, Morselli M, et al. Clinico-pathological characterization of hepatitis C virus-related B-cell non-Hodgkin's lymphomas without symptomatic cryoglobulinemia. *Ann Oncol* 1998;9:495–498.
- [122] Ohsawa M, Shingu N, Miwa H, Yoshihara H, Kubo M, Tsukama H, et al. Risk of non-Hodgkin's lymphoma in patients with hepatitis C virus infection. *Int J Cancer* 1999;80:237–239.
- [123] Brind AM, Watson JP, Burt A, Kestevan P, Wallis J, Proctor SJ, et al. Non-Hodgkin's lymphoma and hepatitis C virus infection. *Leuk Lymphoma* 1996;21:127–130.
- [124] Singer IO, Cumming RL, Hogg RB. Is hepatitis C associated with non-Hodgkin's lymphoma? *Leuk Lymphoma* 1997;26:633–634.
- [125] McColl MD, Singer IO, Tait RC, McNeil IR, Cumming RL, Hogg RB. The role of hepatitis C virus in the aetiology of non-Hodgkin's lymphoma – a regional association? *Leuk Lymphoma* 1997;26:127–130.
- [126] Collier JD, Zanke B, Moore M, Kessler G, Krajden M, Shepherd F, et al. No association between hepatitis C and B-cell lymphoma. *Hepatology* 1999;29:1259–1261.
- [127] Panovska I, Georgievski B, Stojanovic A, Cevreska L, Efremov DG. Low prevalence of chronic hepatitis C virus infection in B-cell non-Hodgkin's lymphoma patients from a population with a high prevalence of healthy hepatitis C virus carriers. *Br J Haematol* 2000;109:249–250.
- [128] Germanidis G, Haioun C, Pourquier J, Gaulard P, Pawlotsky JM, Dhumeaux D, et al. Hepatitis C virus infection in patients with overt B-cell non-Hodgkin's lymphoma in a French center. *Blood* 1999;93:1778–1779.
- [129] Thalen DJ, Raemaekers J, Galama J, Cooreman MP. Absence of hepatitis C virus infection in non-Hodgkin's lymphoma. *Br J Haematol* 1997;96:880–881.
- [130] Perez RG, Gross JB, Witzig TE, Germer J, Persig DH. Lack of association between hepatitis C or hepatitis G infection and non-Hodgkin's lymphoma (abstr). *Gastroenterology* 1997;112:A1356.
- [131] Hanley J, Jarvis L, Simmonds P, Parker A, Ludlam C. HCV and non-Hodgkin lymphoma. *Lancet* 1996;347:1339.
- [132] Ray RB, Lagging LM, Meyer K, Ray R. Hepatitis C virus core protein cooperates with ras and transforms primary rat embryo fibroblasts to tumorigenic phenotype. *J Virol* 1996;70:4438–4443.
- [133] Machida K, Shimodaira S, Sung VM, Cheng K, Lindsay KL, Levine AM, et al. Hepatitis C virus is a potent mutator: enhanced hypermutation of immunoglobulin and oncogenes in HCV-infection (abstr). 9th International Meeting on HCV and Related Viruses. San Diego, CA. July 2002; p 196.
- [134] Hermine O, Lefrere F, Bronowicki JP, Mariette X, Jondeau K, Eclache-Saudreau V, et al. Regression of splenic lymphoma with villous lymphocytes after treatment of hepatitis C virus infection. *N Engl J Med* 2002;347:89–94.
- [135] Lanford RE, Chavez D, Chisari FV, Sureau C. Lack of detection of negative-strand hepatitis C virus RNA in peripheral blood mononuclear cells and other extrahepatic tissues by the highly strand-specific rTth reverse transcriptase PCR. *J Virol* 1995;69:8079–8083.
- [136] Muratori L, Gibellini D, Lenzi M, Cataleta M, Muratori P, Morelli MC, et al. Quantification of hepatitis C virus-infected peripheral blood mononuclear cells by in situ reverse transcriptase-polymerase chain reaction. *Blood* 1996;248:164–171.
- [137] Laskus T, Radkowski M, Wang LF, Jang SJ, Vargas H, Rakela J. Hepatitis C virus quasispecies in patients infected with HIV-1: correlation with extrahepatic viral replication. *Virology* 1998;248:164–171.
- [138] Shimizu YK, Igarashi H, Kanematu T, Fujiwara K, Wong DC, Purcell RH, et al. Sequence analysis of the hepatitis C virus genome recovered from serum, liver and peripheral blood mononuclear cells of infected chimpanzees. *J Virol* 1997;71:5769–5773.
- [139] Lerat H, Rumin S, Habersetzer F, Berby F, Trabaud MA, Trépo C, et al. In vivo tropism of hepatitis C virus genomic sequences in hematopoietic cells: influence of viral load, viral genotype, and cell phenotype. *Blood* 1998;91:3841–3849.
- [140] Westermann J, Pabst R. Distribution of lymphocyte subsets and natural killer cells in the human body. *Clin Investig* 1992;70:539–544.
- [141] Zignego AL, Giannelli F, Marocchi ME, Mazzocca A, Ferri C, Giannini C, et al. T(14;18) translocation in chronic hepatitis C virus infection. *Hepatology* 2000;31:474–479.
- [142] Kitay-Cohen Y, Amiel A, Hilzenrat N, Buskila D, Ashur Y, Fejgin M, et al. Bcl-2 rearrangement in patients with chronic hepatitis C associated with essential mixed cryoglobulinemia type II. *Blood* 2000;96:2910–2912.
- [143] Zuckerman E, Zuckerman T, Sahar D, Streichman S, Attias D, Sabo E, et al. Bcl-2 and immunoglobulin gene rearrangement in patients with hepatitis C virus infection. *Br J Haematol* 2001;112:364–369.
- [144] Zignego AL, Ferri C, Giannelli F, Giannini C, Caini P, Monti M, et al. Prevalence of bcl-2 rearrangement in patients with hepatitis C virus-related mixed cryoglobulinemia with or without B-cell lymphomas. *Ann Intern Med* 2002;137:571–580.
- [145] Monteverde A, Ballare M, Pileri S. Hepatic lymphoid aggregates in chronic hepatitis C and mixed cryoglobulinemia. *Springer Semin Immunopathol* 1997;19:99–110.
- [146] Knight GB, Zhang QX, Agnello V. Bcl-2 over-expression in the absence of t(14;18) (q32;q21) chromosomal translocation in hybridomas and lymphocytes expressing monoclonal IgM from HCV patients (abstr). *Antiviral Ther* 2000;5:C42.
- [147] Casato M, Mecucci C, Agnello V, Fiorilli M, Knight GB, Matteucci C, et al. Regression of lymphoproliferative disorder after treatment for hepatitis C virus infection in a patient with partial trisomy 3. Bcl-2 overexpression, and type II cryoglobulinemia. *Blood* 2002;99:2259–2261.
- [148] Papakonstantinou G, Verbeke C, Hastka J, Bohrer M, Hehlmann R. Bcl-2 expression in non-Hodgkin's lymphomas is not associated

- with bcl-2 gene rearrangements. *Br J Haematol* 2001;113:383–390.
- [149] Chan JK. The new World Health Organization classification of lymphomas: the past, the present and the future. *Hematol Oncol* 2001;19:129–150.
- [150] De Re V, De Vita S, Marzotto A, Rupolo M, Gloghini A, Pivetta B, et al. Sequence analysis of the immunoglobulin antigen receptor of hepatitis C virus-associated non-Hodgkin lymphomas suggests that the malignant cells are derived from the rheumatoid factor-producing cells that occur mainly in type II cryoglobulinemia. *Blood* 2000;96:3578–3584.
- [151] De Vita S, Sacco C, Sansonno D, Gloghini A, Dammacco F, Crovatto M, et al. Characterization of overt B-cell lymphomas in patients with hepatitis C virus infection. *Blood* 1997;90:776–782.
- [152] Franzin F, Efremov DG, Pozzato G, Tulissi P, Batista F, Burrone OR. Clonal B-cell expansions in peripheral blood of HCV-infected patients. *Br J Haematol* 1995;90:548–552.
- [153] Pozzato G, Mazzaro C, Crovatto M, Modolo ML, Ceselli S, Mazzi G, et al. Low-grade malignant lymphoma, hepatitis C virus infection, and mixed cryoglobulinemia. *Blood* 1994;84:3047–3053.
- [154] Zignego AL, Ferri C, Giannini C, La Civita L, Carecchia G, Longombardo G, et al. Hepatitis C virus infection in mixed cryoglobulinemia and B-cell non-Hodgkin's lymphoma: evidence for a pathogenetic role. *Arch Virol* 1997;142:545–555.
- [155] Patriarca F, Silvestri F, Fanin R, Zaja F, Sperotto A, Baccarani M. Long-lasting complete remission of hepatitis C virus (HCV) infection and HCV-associated immunocytoma with alpha-interferon treatment. *Br J Haematol* 2001;112:370–372.
- [156] Hermine O, Lefrere F, Bronowicki JP, Mariette K, Jondeau K, Eclache-Saudreau V, et al. Regression of splenic lymphoma with villous lymphocytes after treatment of hepatitis C virus infection. *N Engl J Med* 2002;347:89–94.
- [157] Ivanovski M, Silvestri F, Pozzato G, Anand S, Mazzaro C, Burrone OR, et al. Somatic hypermutation, clonal diversity, and preferential expression of the VH 51p1/VL kv325 immunoglobulin gene combination in hepatitis C virus-associated immunocytomas. *Blood* 1998;91:2433–2442.
- [158] De Vita S, De Re V, Sansonno D, Sorrentino D, Corte RL, Pivetta B, et al. Gastric mucosa as an additional extrahepatic localization of hepatitis C virus: viral detection in gastric low-grade lymphoma associated with autoimmune disease and in chronic gastritis. *Hepatology* 2000;31:182–189.
- [159] Shirin H, Davidovitz Y, Avni Y, Petchenko P, Krepel Z, Bruck R, et al. Prevalence of hepatitis C virus infection in patients with lymphoproliferative disorders. *Isr Med Assoc J* 2002;4:24–27.
- [160] Ellenrieder V, Weidenbach H, Frickhofen N, Michel D, Prummer O, Klatt S, et al. HCV and HGV in B-cell non-Hodgkin's lymphoma. *J Hepatol* 1998;28:34–39.
- [161] Cucuianu A, Patiu M, Duma M, Bararab C, Soritau O, Bojan A, et al. Hepatitis B and C virus infection in Romanian non-Hodgkin's lymphoma patients. *Br J Haematol* 1999;107:353–356.
- [162] Desenne JJ, Somoza R, Torres MA, Acquatella G, Natlae A, Quijada JG, et al. Hepatitis C virus in patients with non-Hodgkin's lymphoma (abstr). *Ann Oncol* 1996;7:86.
- [163] Sikuler E, Shnaider A, Zilberman D, Hilzenrat N, Shemer-Avni Y, Neumann L, et al. Hepatitis C virus infection and extrahepatic malignancies. *J Clin Gastroenterol* 1997;24:87–89.
- [164] Gasztonyi B, Par A, Szomor A, Nagy A, Kereskai L, Losconczy H, et al. Hepatitis C virus infection and B-cell non-Hodgkin's lymphoma. *Orv Hetil* 2000;141:2649–2651.
- [165] Hausfater P, Cacoub P, Sterkers Y, Thibault V, Amoura Z, Nguyen L, et al. Hepatitis C virus infection and lymphoproliferative diseases: prospective study on 1576 patients in France. *Am J Hematol* 2001;67:168–171.
- [166] Zuckerman T, Zuckerman E, Douer D, Fong TL, Nathwani BN, Velankar M, et al. Hepatitis C virus infection in patients with B-cell non-Hodgkin's lymphoma (abstr). *Blood* 1996;88:887.
- [167] Mangia A, Clemente R, Musto P, Cascavilla I, La Floresta P, Sarpaolo G, et al. Hepatitis C virus infection and monoclonal gammopathies not associated with cryoglobulinemia. *Leukemia* 1996;10:1209–1213.
- [168] Cavanna L, Sbolli G, Tanzi E, Romano L, Civardi G, Buscarini E, et al. High prevalence of antibodies to Hepatitis C Virus in patients with lymphoproliferative disorders. *Haematologica* 1995;80:486–487.
- [169] Thalen D, Raemaekers J, Galama J, Cooreman M. Non-Hodgkin's lymphoma is not related to hepatitis C virus infection (abstr). *Ann Oncol* 1996;7:86.
- [170] Tkoub EM, Haioun C, Pawlotsky JM, Dhumeaux D, Delchier JC. Chronic hepatitis C virus and gastric MALT lymphoma. *Blood* 1998;91:360.
- [171] Ohsawa M, Shingu N, Miwa H, Yoshihara H, Kubo M, Tsukuma H, et al. Risk of non-Hodgkin's lymphoma in patients with hepatitis C virus infection. *Int J Cancer* 1999;80:237–239.
- [172] Shariff S, Yoshida EM, Gascoyne RD, Le N, Connors JM, Middleton PJ, et al. Hepatitis C infection and B-cell non-Hodgkin's lymphoma in British Columbia: a cross-sectional analysis. *Ann Oncol* 1999;10:961–964.
- [173] King PD, Wilkes JD, Diaz-Arias AA. Hepatitis C virus infection in non-Hodgkin's lymphoma. *Clin Lab Haematol* 1998;20:107–110.
- [174] Udomsakdi-Auewarakul C, Auewarakul P, Sukpanichnant S, Muangsup W. Hepatitis C virus infection in patients with non-Hodgkin lymphoma in Thailand. *Blood* 2000;85:3640–3641.
- [175] Ferri C, Caracciolo F, Zignego AL, La Civita L, Monti M, Longombardo G. Hepatitis C virus infection in patients with non-Hodgkin's lymphoma. *Br J Haematol* 1994;88:392–394.
- [176] Pioltelli P, Gargantini L, Cassi E, Santoleri L, Bellati G, Magliano EM, et al. Hepatitis C virus in non-Hodgkin's lymphoma. A reappraisal after a prospective case-control study of 300 patients. *Lombard Study Group of HCV-Lymphoma. Am J Hematol* 2000;64:95–100.
- [177] Mazzaro C, Zagonel V, Monfardini S, Tulissi P, Pussini E, Fanni M, et al. Hepatitis C virus and non-Hodgkin's lymphomas. *Br J Haematol* 1996;94:544–550.
- [178] De Rosa G, Gobbo ML, De Renzo A, Notaro R, Garofalo S, Grimaldi M. High prevalence of hepatitis C virus infection in patients with B-cell lymphoproliferative disorders in Italy. *Am J Hematol* 1997;55:77–82.
- [179] Hezode C, Duvoux C, Germanidis G, Roudot-Thoraval F, Vincens AL, Gaullard P, et al. Role of hepatitis C virus in lymphoproliferative disorders after liver transplantation. *Hepatology* 1999;30:775–778.
- [180] Viguier M, Rivet J, Agbalika F, Kerviler E, Brice P, Dubertret L, et al. B-cell lymphomas involving the skin associated with hepatitis C virus infection. *Int J Dermatol* 2002;41:577–582.
- [181] Chowla A, Malhi-Chowla N, Chidambaram A, Surick B. Primary hepatic lymphoma in hepatitis C: case report and review of the literature. *Am Surg* 1999;65:881–883.
- [182] De Vita S, De Re V, Gasparotto D, Ballare M, Pivetta B, Ferraccioli G, et al. Oligoclonal non-neoplastic B cell expansion is the key feature of type II mixed cryoglobulinemia: clinical and molecular findings do not support a bone marrow pathologic diagnosis of indolent B cell lymphoma. *Arthritis Rheum* 2000;43:94–102.
- [183] Mele A, Pulsoni A, Bianco E, Musto P, Szklo A, Sanpaolo MG, et al. Hepatitis C virus and B-cell non-Hodgkin lymphomas: an Italian multicenter case-control study. *Blood* 2003;102:996–999.
- [184] Limpens J, Stad R, Vos C, de Vlaam C, de Jong D, van Ommen GJ, et al. Lymphoma-associated translocation t(14;18) in blood B cells of normal individuals. *Blood* 1995;85:2528–2536.
- [185] Dolken G, Illerhaus G, Hirt C, Mertelsmann R. BCL-2/JH rearrangements in circulating B cells of healthy blood donors and patients with non-malignant diseases. *J Clin Oncol* 1996;14:1333–1344.
- [186] Summers KE, Goff LK, Wilson AG, Gupta RK, Lister TA, Fitzgibbon J. Frequency of the Bcl-2/IgH rearrangement in normal

- individuals: implications for the monitoring of disease in patients with follicular lymphoma. *J Clin Oncol* 2001;19:420–424.
- [187] Liu Y, Hernandez AM, Shibata D, Cortopassi GA. BCL2 translocation frequency rises with age in humans. *Proc Natl Acad Sci USA* 1994;91:8910–8914.
- [188] Poetsch M, Weber-Matthiesen K, Plendl HJ, Grote W, Schlegelberger B. Detection of the *t*(14;18) chromosomal translocation by interphase cytogenetics with yeast-artificial-chromosome probes in follicular lymphoma and non-neoplastic lymphoproliferation. *J Clin Oncol* 1996;14:963–969.
- [189] clinical A. evaluation of the International Lymphoma Study Group classification of non-Hodgkin's lymphoma. The Non-Hodgkin's Lymphoma Classification Project. *Blood* 1997;89:3909–3918.
- [190] De Vita S, Zagonel V, Russo A, Rupolo M, Cannizzaro R, Chiara G, et al. Hepatitis C virus, non-Hodgkin's lymphomas and hepatocellular carcinoma. *Br J Cancer* 1998;77:2032–2035.
- [191] Spencer J, Perry ME, Dunn-Walters DK. Human marginal-zone B cells. *Immunol Today* 1998;19:421–426.
- [192] Sansonno D, De Vita S, Iacobelli AR, Cornacchiulo V, Boiocchi M, Dammacco F. Clonal analysis of intrahepatic B cells from HCV-infected patients with and without mixed cryoglobulinemia. *J Immunol* 1998;160:3594–3601.
- [193] Knight G, Agnello V. WA monoclonal rheumatic factors and non-Hodgkin Lymphoma. *Blood* 2001;97:3319–3321.
- [194] Chan CH, Hadlock KG, Fong SK, Levy S. V(H)1-69 gene is preferentially used by hepatitis C virus-associated B cell lymphomas and by normal B cells responding to the E2 viral antigen. *Blood* 2001;97:1023–1026.
- [195] Quinn ER, Chan CH, Hadlock KG, Fong SK, Flint M, Levy S. The B-cell receptor of a hepatitis C virus (HCV)-associated non-Hodgkin lymphoma binds the viral E2 envelope protein, implicating HCV in lymphomagenesis. *Blood* 2001;98:3745–3749.